A diabetic with high haemoglobin A1c due to persistent haemoglobin F

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Abstract

Laboratory and patient-related factors can result in false glycated haemoglobin (HbA1c) measurements. Haemoglobin (Hb) variants that interfere with laboratory readings is an important cause. We report a case of hereditary persistence of Fetal Haemoglobin manifesting as a falsely high HbA1c in a 35-year old patient with type 2 diabetes mellitus, whose high HbA1c values persisted despite intensive anti-diabetic treatment. His fasting and postprandial blood glucose values as well as serum fructosamine level was incongruously low compared to HbA1c values. The presence of fetal haemoglobin was confirmed by haemoglobin electrophoresis. This case highlights the importance of being aware of the factors that can influence laboratory HbA1c measurements.

Introduction

Glycated haemoglobin (HbA1c) is a widely used measure of glycaemic control. Haemoglobin (Hb) variants that interfere with laboratory readings is an important cause. We present a case of an Hb variant causing aberrantly high HbA1c values in a patient with diabetes, and review some of the factors that affect HbA1c measurements.

Case report

A 35-year old welder was referred to the endocrinologist by a general practitioner for the management and follow up of very poorly controlled diabetes mellitus. He had presented one month ago with a fainting episode elevated capillary blood glucose. He was started on Metformin and Glibenclamide after the confirmation of diabetes and later referred to the endocrinologist as his HbA1c was 70%. He did not have a significant past medical history and had never required blood transfusions. However, he had a strong family history of diabetes, with both his parents and two other siblings affected from their early fifties. On examination he was an averagely built man who was not pale or icteric. He had a 2 cm hepatomegaly and a 4 cm splenomegaly. The rest of the clinical examination was unremarkable with no evidence of diabetic retinopathy or neuropathy.

His fasting blood glucose was 154 mg/dl and the post prandial blood glucose was 224 mg/dl. However, his HbA1c was 70% which was discrepantly high compared to his blood glucose values. The HbA1c test was done by ion exchange high performance liquid chromatography (HPLC) using a Biorad D10 machine. The possibility of an erroneous HbA1c reading was considered. This was supported by a fructosamine level of 294 micmol/l (205-285 micmol/l), which denoted only a slightly impaired glucose control. Since Hb variants are known to cause aberrantly high HbA1c values during laboratory testing, we considered a Hb variant to be a strong possibility in our patient who had hepatosplenomegaly. His Hb count was 15.2 g/dl and the blood picture showed hypochromic microcytic red cells, numerous target cells, irregularly contracted cells and irregularly haemoglobinized cells suggesting a thalassaemia trait. His haemoglobin electrophoresis revealed that his haemoglobin consisted entirely of Hb F (foetal haemoglobin) with no detectable Hb A or HbA2. This was suggestive of hereditary persistence of foetal haemoglobin. Although he was a product of a consanguineous marriage a similar illness had not been diagnosed in any of his family members previously.

Discussion

This is an interesting presentation of an otherwise undetected hereditary persistence of haemoglobin F manifesting solely as a discrepantly high HbA1c value.

In healthy adults haemoglobin comprises 97% of Hb A, 2.5% of HbA2 and 0.5% of HbF. Fractionation of Hb A by chromatography identifies several minor peaks referred to as Hb A1a or fast Hbs which include HbA1a, HbA1b and HbA1c. Glucose binds to haemoglobin in a two step process, and as one is irreversible, once bound

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it lasts through the lifespan of the red blood cell, approximately 2 to 3 months. The N terminal valine of beta chains provides the most common site of glycation within the haemoglobin tetramer, accounting for 80% of HbA1c (1).

Hence HbA1c is the most widely used to monitor glycaemic control during a period of approximately 3 months and strongly correlates with the mean blood glucose level. The Diabetes Control and Complications trial (DCCT) and the United Kingdom Prospective Diabetes Study (UKPDS) were large trials that demonstrated that HbA1c levels are directly related to the risk of complications of diabetes, stroke and ischaemic heart disease (2, 3). As HbA1c is also a strong predictor of new onset diabetes mellitus, the American Diabetes Association now recommends this test in the diagnosis of diabetes mellitus and for identifying pre-diabetes (4). The American College of Endocrinology (ACE) and American Association of Clinical Endocrinologists (AACE) perceive that HbA1c should not be the primary criterion for the diagnosis of diabetes mellitus and that it should be used in conjunction with fasting plasma glucose and/or oral glucose tolerance tests (5).

As HbA1c is important as a diagnostic and monitoring tool, it is important to be aware of conditions that can affect laboratory HbA1c values apart from plasma glucose levels. Conditions that cause increased cell turnover and reduced average life span of the red blood cells can lead to lower HbA1c values. These include active bleeding, haemolytic disease, haemoglobinopathies and myelodysplastic disease. A patient with renal failure and uraemia can have high concentrations of carbamylated haemoglobin, resulting in aberrantly high HbA1C. Falsely elevated HbA1c measurements may also be obtained when red blood cell turnover is low, resulting in higher proportions of older red blood cells, such as in iron, B12 or folate deficiencies. Haemoglobinopathies can affect HbA1c values in three ways, they can influence the binding of glucose to haemoglobin, affect the peak measurements on chromatography and increase the risk of haemolysis and hence decrease the lifespan of red blood cells (6).

There are several commonly available methods to calculate HbA1c such as cation exchange chromatography, boronate affinity chromatography and immunoassay methods. Glycation alters the structure of the haemoglobin molecule and decreases its positive charge. Cation exchange chromatography separates haemoglobin species based on charge difference. Hb species are eluted from the cation exchange column at different times with the application of buffers of increasing ionic strength. A spectrophotometer measures the concentration of Hb in each column which is then quantified by calculating the area under each peak. The HbA1c percentage is then determined by an equation which includes HbA and HbA1c values (1). Ion exchange high performance liquid chromatography (HPLC) method, which was the method used for our patient by the Biorad D10 machine uses similar principles.

Carriers of variant Hbs that elute separately from HbA and HbA1c theoretically should have little effect on the HbA1c measurement as they have little effect on the equation. However, several reports have indicated that Hb variants and elevated HbF levels can interfere with some HbA1c assays (1). However, only a few studies are available. One such study showed that ion exchange HPLC methods show only very minimal evidence of interference from elevated HbF levels even when the HbF levels exceed 15% (7). As our patient had 100% of HbF it probably interfered with the spectrophotometry of haemoglobin columns and gave the erroneous HbA1c value. Although it was previously thought that the boronate affinity method was not affected by haemoglobin variants, the previously mentioned study showed that the presence of HbF artificially lowered the HbA1c values by this method as well (7). International Federation of Clinical Chemistry (IFCC) Reference Method (IFCC RM) for HbA1c measures glycated and nonglycated hexapeptides from HbA β chains. Because HbF has no β chains, HbF does not cause interference with the IFCC RM because only the HbA terminal hexapeptides are measured. Therefore this method has minimal interference by HbF and can be used to assess glycaemic control in these patients (8).

When there are inconsistencies between a patient’s home blood glucose monitoring and laboratory measured HbA1c, one should suspect a falsely elevated or lowered HbA1c result. Suspicion should also be raised when HbA1c is more than 15%, or when there is a significant change in a patient’s HbA1c when the laboratory assay method is changed (9). Comparing the patients home blood glucose monitoring values with his venous plasma glucose values would verify the accuracy of the blood glucose readings as opposed to the HbA1c value indicating the need to further evaluate a reason for the discrepantly high HbA1c value. Serum fructosamine level which reflects the average blood glucose control within the previous 2 to 3 weeks could also be used as a surrogate test. However the correlation between fructosamine and complications of diabetes mellitus has not been robustly evaluated in large randomised trials.

Elevated HbF levels can occur in patients as a result of pathologic conditions (eg, thalassaemia and leukaemia) or hereditary persistence of fetal haemoglobin (1). Approximately 1.5% of the US population has been reported to have elevated HbF levels as defined by an HbF level of more than 2% (10). Patients with the most common form of hereditary persistence of fetal haemoglobin can have HbF levels of up to 30%, and because they are generally asymptomatic, patients and their physicians may be unaware of the existence of this
condition (10). Our patient had 100% of HbF but was otherwise asymptomatic making hereditary persistence of foetal haemoglobin the most likely possibility in our patient.

In summary, this case illustrates the importance of maintaining a high degree of suspicion when the blood glucose values in a diabetic are incongruous with the HbA1c values. It also emphasizes the importance of using other biochemical parameters to diagnose and monitor glycaemic control in patients with haemoglobinopathies and haemoglobin variants.

References


